

Scotland's Rural College

The contribution of cattle urine and dung to nitrous oxide emissions: quantification of country specific emission factors and implications for national inventories

Chadwick, DR; Cardenas, LM; Dhanoa, MS; Donovan, N; Misselbrook, T; Williams, JR; Thorman, RE; McGeough, KL; Watson, CJ; Bell, M; Anthony, SG; Rees, RM

Published in:
Science of the Total Environment

DOI:
[10.1016/j.scitotenv.2018.04.152](https://doi.org/10.1016/j.scitotenv.2018.04.152)

Print publication: 01/09/2018

Document Version
Publisher's PDF, also known as Version of record

[Link to publication](#)

Citation for pulished version (APA):

Chadwick, DR., Cardenas, LM., Dhanoa, MS., Donovan, N., Misselbrook, T., Williams, JR., Thorman, RE., McGeough, KL., Watson, CJ., Bell, M., Anthony, SG., & Rees, RM. (2018). The contribution of cattle urine and dung to nitrous oxide emissions: quantification of country specific emission factors and implications for national inventories. *Science of the Total Environment*, 635, 607 - 617. <https://doi.org/10.1016/j.scitotenv.2018.04.152>

General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal ?

Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Download date: 05. May. 2023



The contribution of cattle urine and dung to nitrous oxide emissions: Quantification of country specific emission factors and implications for national inventories

D.R. Chadwick^{a,*}, L.M. Cardenas^b, M.S. Dhanoa^b, N. Donovan^b, T. Misselbrook^b, J.R. Williams^c, R.E. Thorman^c, K.L. McGeough^d, C.J. Watson^d, M. Bell^e, S.G. Anthony^f, R.M. Rees^e

^a School of Environment, Natural Resources and Geography, Bangor University, Bangor LL57 2UW, UK

^b Rothamsted Research, North Wyke, Devon EX20 2SB, UK

^c ADAS Boxworth, Battlegate Rd., Cambridge CB23 4NN, UK

^d Agri-Food and Biosciences Institute, 18a, Newforge Lane, BT9 5PX, Belfast, UK

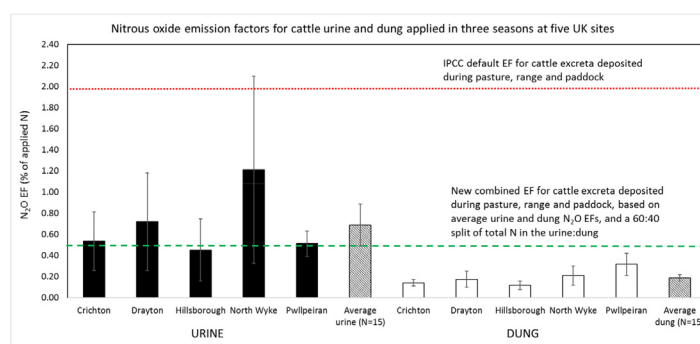
^e Scotland's Rural College (SRUC), West Mains Road, Edinburgh EH9 3JG, UK

^f ADAS Wolverhampton, Titan 1 offices, Coxwell Avenue, Wolverhampton Science Park, Wolverhampton WV10 9RT, UK

HIGHLIGHTS

- First co-ordinated experiments in UK to generate data for country specific grazing excretal N₂O EF
- Urine had a significantly greater average N₂O EF (0.69%) than dung (0.19%).
- The combined excretal N₂O EF was 0.49%, <25% of the IPCC default value for cattle.
- DCD reduced the N₂O EF from urine patches by an average of 46%.
- Urine N₂O was controlled by its composition, dung N₂O was related to soil and environmental factors.

GRAPHICAL ABSTRACT



ARTICLE INFO

Article history:

Received 12 March 2018

Received in revised form 10 April 2018

Accepted 10 April 2018

Available online 24 April 2018

Editor: Jay Gan

Keywords:

Grassland
Greenhouse gas
Nitrous oxide
Cattle
Urine patch
Dung pat
Nitrification inhibitor

ABSTRACT

Urine patches and dung pats from grazing livestock create hotspots for production and emission of the greenhouse gas, nitrous oxide (N₂O), and represent a large proportion of total N₂O emissions in many national agricultural greenhouse gas inventories. As such, there is much interest in developing country specific N₂O emission factors (EFs) for excretal nitrogen (EF₃, pasture, range and paddock) deposited during grazing. The aims of this study were to generate separate N₂O emissions data for cattle derived urine and dung, to provide an evidence base for the generation of a country specific EF for the UK from this nitrogen source. The experiments were also designed to determine the effects of site and timing of application on emissions, and the efficacy of the nitrification inhibitor, dicyandiamide (DCD) on N₂O losses. This co-ordinated set of 15 plot-scale, year-long field experiments using static chambers was conducted at five grassland sites, typical of the soil and climatic zones of grazed grassland in the UK. We show that the average urine and dung N₂O EFs were 0.69% and 0.19%, respectively, resulting in a combined excretal N₂O EF (EF₃), of 0.49%, which is <25% of the IPCC default EF₃ for excretal returns from grazing cattle. Regression analysis suggests that urine N₂O EFs were controlled more by composition than was the case for dung, whilst dung N₂O EFs were more related to soil and environmental factors. The urine N₂O EF was significantly greater from the site in SW England, and significantly greater from the early grazing

* Corresponding author.

E-mail address: d.chadwick@bangor.ac.uk (D.R. Chadwick).

season urine application than later applications. Dicyandiamide reduced the N_2O EF from urine patches by an average of 46%. The significantly lower excretal EF₃ than the IPCC default has implications for the UK's national inventory and for subsequent carbon footprinting of UK ruminant livestock products.

© 2018 The Authors. Published by Elsevier B.V. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Grazed grasslands support a significant proportion of sheep and cattle production throughout Europe and other parts of the World, converting human-inedible plant biomass into human edible animal products but with generally low nitrogen (N) use efficiencies. The ruminant animal converts much of the organic N in plant biomass into highly reactive and bioavailable N (Nr), particularly as excreted in the urine. It is thought that 3.08 Mt. of N is deposited by grazing livestock in Europe, and this value is thought to be as much as ca. 0.61 Mt. N in the UK (UNFCCC, 2016). It is well documented that urine additions to grassland soils result in significant quantities of N_2O production and emission, mainly due to the soil microbial processes of nitrification and denitrification (Selbie et al., 2015), following the addition of readily available N and carbon (C), and the effects of significantly increased percentage of water-filled pore space (WFPS) within the urine patch (van der Weerden et al., 2017).

Deposition of N in urine patches can represent an equivalent application rate of 200–2000 kg N ha⁻¹ (Selbie et al., 2015), depending on the protein content of the sward, livestock type, age and stage of lactation. A meta-analysis by Selbie et al. (2015) indicates average urine patch N loading rates of 613 kg N ha⁻¹ and 345 kg N ha⁻¹ for dairy cows and beef cattle, respectively. Clearly, N loading rates in urine patches are in excess of optimal plant use efficiency, increasing the risk of excess N being lost to the environment via nitrate (NO_3^-) leaching (de Klein and Ledgard, 2001; Di and Cameron, 2007), ammonia (NH_3) volatilization (Lockyer and Whitehead, 1990; Laubach et al., 2013; Burchill et al., 2017), N_2O (Di and Cameron, 2008; Krol et al., 2016; Van der Weerden et al., 2017; Minet et al., 2018) and N_2 (Clough et al., 1998) emissions. All of these N loss pathways (except N_2O losses) typically represent a significant agronomic loss, and all but N_2 loss have detrimental effects on the environment.

At these high rates of N loading, the N_2O emission is likely to be disproportionately greater than emissions from N sources applied at lower N loading rates, e.g. typical fertiliser N applications at agronomic rates. A curvilinear response of N_2O emissions to N loading has been shown previously, e.g. Cardenas et al. (2010) for fertiliser N (NH_4NO_3) applications between 0 and 375 kg N ha⁻¹ to grazed swards. Bell et al. (2015) also showed a non-linear response of N_2O fluxes to NH_4NO_3 applications (0–400 kg N ha⁻¹) to cut grass. More specifically for urine applications, de Klein et al. (2014) demonstrated greater N_2O emissions, as a percentage of N applied, i.e. emission factors (EFs), (0.34%) from urine patches receiving an N loading of 1200 kg ha⁻¹ compared to urine patches with a lower N loading (0.10% from a loading of 200 kg ha⁻¹) on a freely draining soil, although a linear relationship between N_2O EFs and urine N loading was observed on a poorly drained soil. van Groenigen et al. (2005) found no effect of N loading in urine patches on the N_2O EF.

For excretion during cattle grazing, the default IPCC N_2O EF (pasture, range and paddock) is 2% for (combined excretal urine + dung EF) (cf to 1% for fertiliser N), whilst the N_2O EF for sheep excretal N during grazing is only 1% (IPCC, 2006). UNFCCC submissions for 2015 from different countries (using IPCC Tier 1/2, 2006 Guidelines) show that direct N_2O emissions following N deposited to soil by grazing livestock represents from <5% (e.g. in Japan) to >65% (in New Zealand) of total national direct soil N_2O emissions (Fig. 1), with greater contributions coming from countries where livestock graze for significant periods of the year (UNFCCC, 2016). As this source of direct N_2O emissions is significant to many national agricultural greenhouse gas inventories, there is

increasing interest in developing country specific EFs that better reflect national soils and climatic conditions (e.g. Krol et al., 2016 for Ireland).

Most N_r excreted during grazing is in the urine, which is mostly comprised of urea that requires hydrolysis to free NH_4^+ (Selbie et al., 2015). In dung, most N is in the organic form, and requires mineralisation over a longer time period to provide a pool of NH_4^+ for nitrification and NO_3^- for denitrification. The split between urine and dung for total excretal N will depend on dietary protein intake compared with requirement by the animal (as protein intake increases above requirement proportionally more N will be excreted as urine (Broderick, 2003; Reed et al., 2015), and partially on the digestibility of the protein in the diet (with a higher proportion of less digestible protein being excreted as faecal N). The UK GHG and ammonia emission inventories to date have assumed 60% of total N excretion by cattle to be as urine and 40% as dung (Webb and Misselbrook, 2004), in common with other Western European countries (Reidy et al., 2008). Disaggregating emissions to urine and dung offers an improved understanding of the sources of N_2O from grazed pastures, and hence how they could be mitigated.

Since direct N_2O emissions from grazing livestock represent such a large term in national agricultural greenhouse gas inventories, there has been significant interest in understanding factors that contribute to N_2O production and emission from this source, e.g. soil type (Clough et al., 1998), urine composition (Kool et al., 2006; Gardiner et al., 2016), weather conditions (Krol et al., 2016), and in exploring strategies to reduce emissions. For example, Monaghan and de Klein (2014) have suggested restricting the duration of autumn and winter grazing to reduce higher N_2O fluxes associated with urine deposition to wet soils (Qui et al., 2010; Krol et al., 2016). Other studies have explored how manipulating the natural urine composition, e.g. hippuric acid content, can reduce N_2O production from the urine patch (Clough et al., 2009), and there has been much interest in the use of synthetic nitrification inhibitors to reduce both NO_3^- leaching and N_2O emissions from urine patches (Hatch et al., 2005; Di and Cameron, 2012; Barneze et al., 2015). New Zealand and Irish research groups have taken this a step further, in exploring how the nitrification inhibitor dicyandiamide (DCD) can be delivered to urine patches to reduce N_2O emissions, e.g. through boluses (Ledgard et al., 2008), in drinking water (Welten et al., 2014), and in feed (Luo et al., 2015; Minet et al., 2016, 2018). However, recent publicity and research has demonstrated that there are potential unintended consequences of using nitrification inhibitors, such as contamination of milk products, e.g. via root or foliar uptake (Marsden et al., 2015; Pal et al., 2016) and increased ammonia emissions (Lam et al., 2017), so researchers are exploring new inhibitor products, including biological nitrification inhibitory compounds targeted at ruminant production (Gardiner et al., 2016; Balvert et al., 2017; Luo et al., 2018) that may be deemed more acceptable to the public in the future.

The UK greenhouse gas R&D community undertook a large number of field trials to quantify N_2O EFs from a range of different N sources (viz, different fertiliser N forms, different manure types, and urine and dung deposited by grazing livestock (Chadwick et al., 2011), as part of a larger programme to improve the reporting tool for the national inventory of agricultural greenhouse gas emissions that better represents the soils, climate and N management in the UK. In this paper, we summarise the results of the first co-ordinated set of plot-based experiments aimed at generating new N_2O emissions data for disaggregated urine and dung deposition to soil, from which country specific N_2O EFs can be derived that are relevant to UK soils and climate. Some of

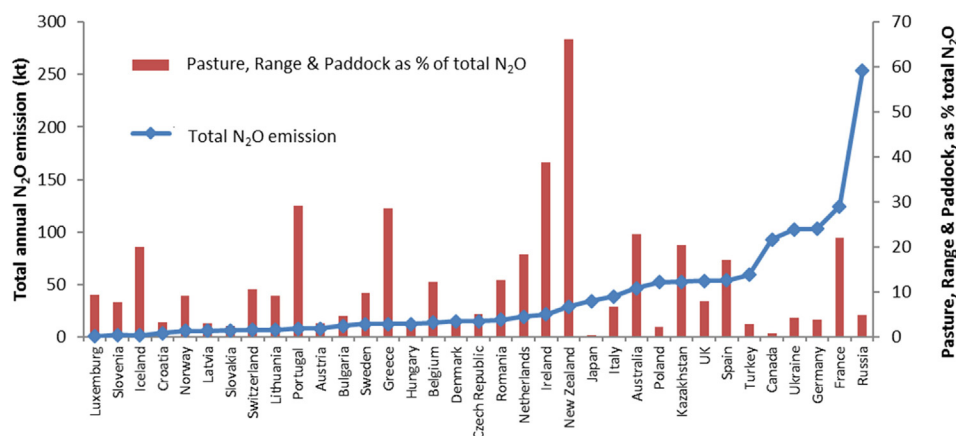


Fig. 1. Annual (year 2014) agricultural N₂O emissions and direct N₂O emissions from excreta deposited by grazing livestock (pasture, range and paddock), expressed as a percentage of the total agricultural N₂O emission, from different nations (source: UNFCCC, 2016).

the individual site experimental results can be found in Bell et al. (2015) and Cardenas et al. (2016). In the experiments, we tested whether season of urine and dung deposition (early grazing, mid grazing, later grazing period) influenced the N₂O EF. We also tested the efficacy of the nitrification inhibitor, dicyandiamide (DCD), to reduce N₂O emissions. An additional reference treatment was included in each experiment, a standardised artificial (synthetic, produced in the laboratory) urine treatment, with the aim of using the information from this treatment to help disentangle the effects of urine composition from soil and climate effects on N₂O EFs.

The specific aims of this study were to: i) determine separate direct N₂O EFs for cattle urine and dung, ii) determine if season of urine and dung deposition affected the direct N₂O emission, iii) assess the effects of site on direct N₂O emissions from urine, iv) evaluate the efficacy of the nitrification inhibitor, DCD, to reduce direct N₂O emissions from urine, and v) assess the influence of using the combined experimentally derived urine and dung N₂O EF on national N₂O emissions.

2. Materials and methods

2.1. Site selection

Five experimental sites were selected to cover the range of typical grassland soils and climate throughout the UK, with two sites in England, one in Scotland, one in Wales and one in Northern Ireland (see locations in Fig. 2). Descriptions of the sites are shown in Table 1. There have been few previous studies in the UK where N₂O EFs have been quantified from urine and dung deposition that are IPCC compliant (IPCC, 2000, 2006) (i.e. where emission measurements were also made from control plots, and where measurements lasted for up to 365 days), that these sites needed to provide an appropriate range of

soil texture and climate. However, some practicality was also considered in site selection; location could not be excessively far from a research base to ensure timely measurements, since >30 measurement occasions were needed during each 12-month experimental period. Four measurement teams, from different UK organisations, ADAS, AFBI, Rothamsted Research – North Wyke and SRUC, conducted the 15 experiments, following an agreed joint experimental protocol to ensure aspects of the urine and dung management, chamber deployment, and ancillary measurements were made in a similar way.

Experiments were conducted on established grasslands where the dominant pasture plant was *Lolium perenne*, which is typical of UK livestock systems (Fig. 2). Each experiment comprised three replicate blocks with five treatments, so a total of 15 plots were sampled on every occasion. There were 5 urine patches or 5 dung pats per plot (to account for variability in soil conditions) with one chamber per patch/pat, hence 45 chambers per experiment. There were also control plots that received no treatment application. Applications were made in the spring, summer and autumn (to separate plots), to simulate excretal deposition in early-, mid- and late- grazing season. Livestock were excluded from grazing the experimental areas at least 6 months prior to the start of any experiment. This minimised any direct effect of previous deposition of excreta on N₂O emissions.

2.2. Urine and dung provision

The experimental design resulted in the need for ca. 200 l of fresh cattle urine and ca. 300 kg dung for each experiment. Urine and dung were collected from the institutions summarised in Table 2 within 7 days of an experiment starting, and stored in sealed containers (unacidified) at <4 °C. Table 2 summarises the origin of the urine and dung used in each experiment.

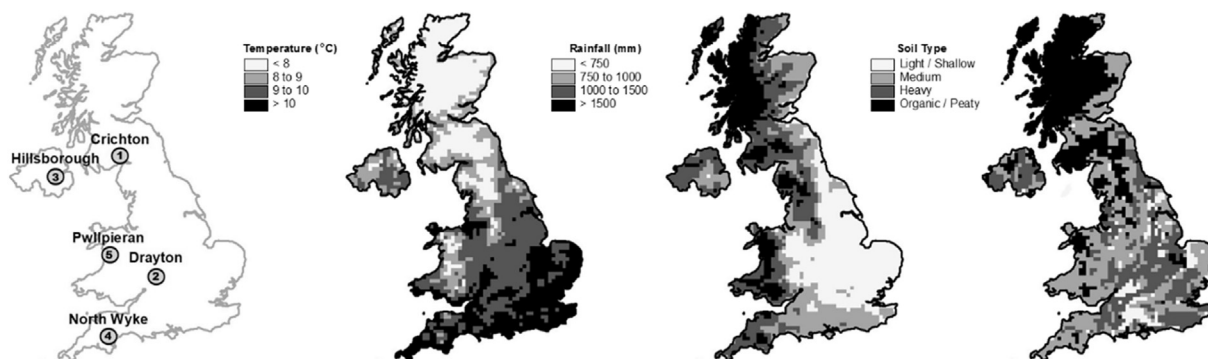


Fig. 2. Site location, climate average (1981 to 2010) rainfall and temperature, and distribution of dominant soil types.

Table 1

Site and soil characteristics. Soil parameters for the 0–10 cm layer.

Site	Country	Altitude (m)	30 yr average annual rainfall (mm)	30 yr average annual air temperature (°C)	Clay content (%)	Soil pH	Organic matter content (%)	Bulk density (g cm ⁻³)
Crichton	Scotland	50	1140	9.1	13	5.6	3.05	1.07
Drayton	England	47	628	10.3	59	7.6	4.84	0.90
Hillsborough	Northern Ireland	128	908	9.0	23	6.0	9.82	0.90
North Wyke	England	185	1042	10.0	37	5.7	5.40	0.62
Pwllpeiran	Wales	213	1570	10.0	29	5.5	5.40	0.92

2.3. Treatments

Urine and dung were removed from cold storage at least 12 h before application to the soil, to allow them to attain ambient temperature prior to application to the soil. Urine and dung were applied at typical N loading rates and volumes. The volumetric loading rate was based on a typical 1.8 l per urination event (Misselbrook et al., 2016). Since the N content of the collected urine varied between feeding trials, the N loading rate varied between an equivalent rate of 340 and 570 kg ha⁻¹, with an average loading rate of 455 kg N ha⁻¹ (see Table 4a). Dung was applied at an equivalent rate of 20 kg m⁻², representing typical deposition by grazing cattle (Sugimoto and Ball, 1989), with an average loading rate of 835 kg N ha⁻¹ (range 625–1020 kg N ha⁻¹; Table 4b). Since urine composition could not be controlled between experiments, a standard artificial urine treatment was included at each site as a reference treatment. This was to allow the effects of soil and climate to be determined. The artificial urine recipe of Kool et al. (2006) was used in all experiments.

A urine treatment containing DCD was added, with DCD applied at a rate of 10 kg ha⁻¹ equivalent (supplying 6.5 kg N ha⁻¹ equivalent), and was mixed with urine (only) just before application, to maximise initial co-location of DCD and NH₄⁺ in the soil profile. This approach also simulated the effect of delivering DCD via boluses (Ledgard et al., 2008), feed (Luo et al., 2015; Minet et al., 2016, 2018) and via water troughs (Welten et al., 2014). The following treatments were established:

- Urine (target 500 kg N ha⁻¹)
- Urine + DCD (target 500 kg N ha⁻¹ + 6.5 kg N ha⁻¹ in DCD)
- Artificial urine (500 kg N ha⁻¹; Kool et al., 2006 recipe)
- Dung (target 800 kg N ha⁻¹)
- Control (no additions)

Five chambers were set up for each treatment plot, and three replicate plots per treatment were arranged in three blocks. Tables 4a and 4b shows application rates for urine and dung at each site.

2.4. Treatment applications

Urine treatments were applied to an area of 0.6 m × 0.6 m within a frame to facilitate infiltration (rather than runoff) using a watering

can. After application, static chambers were inserted centrally into this area. Dung pats were spread to cover the entire area within the chamber. We recognise that urine and dung patches are not normally this large, and have “edges”, but this method of application was deemed the most appropriate to simulate the urine patch and dung pat. It is possible that by applying the N source across the whole area of the chamber that N₂O production and emission may have been affected, but there is no evidence to suggest that this would result in either an under- or over-estimate of the true emission (Marsden et al., 2016). In addition to the urine and dung patches that were established for the N₂O chamber measurements, larger areas of grassland (2 m × 2 m) on each plot (i.e. three replicates per treatment) were treated with either urine or dung at the same rate, allowing multiple soil sampling occasions for soil NO₃⁻, soil NH₄⁺ and soil moisture.

2.5. Nitrous oxide measurements

We used the non-steady state static chamber approach to measure N₂O fluxes (Cardenas et al., 2016). The shape and size of the chambers were 0.4 m × 0.4 m × 0.25 m (high) for the ADAS, North Wyke and AFBI experiments, and 0.4 m diameter × 0.3 m (high) for the SRUC experiments, with individual chamber areas of 0.16 and 0.13 m², respectively. Chambers were opaque. Chamber headspace sampling followed the protocol detailed in Chadwick et al. (2014), whereby chambers were closed for a period of 40 min and a headspace sample taken at this time (T₄₀). Ten ambient air samples (5 at the start and 5 at the end of the chamber closure period) were used to provide the T₀ concentration. Gas samples were placed in pre-evacuated 20 ml vials and transported back to individual laboratories for analysis by gas chromatography. Five chambers were assigned randomly per plot; these generated one mean flux per plot. The headspace sampling assumed a linear increase in headspace N₂O concentration (as evidenced by previous research; Chadwick et al., 2014). This linear response was checked on each sampling occasion by measuring the headspace concentration at 10 min intervals up to 60 min after closure, from one chamber per block.

Sampling frequency was 4–5 times in the first week after treatment application, 4–5 times in the second week, 2 times per week for the next two weeks, then once per week for 1 month. Sampling frequency was then reduced further, eventually to once per month until the end of the experiment (12 months), resulting in ca. 30 samples over the 12-

Table 2

Sources of urine and dung for the experiments.

	Cattle type	Age and approx. live weight	Diet
Crichton	Lactating dairy cows	3–7 years old (ca 600 kg)	Grass silage + concentrates (6.5 kg DM head ⁻¹ day ⁻¹)
Drayton	Lactating dairy cows	6 years old (ca. 600 kg)	Concentrate blend, hay, straw, grass silage, maize silage
Hillsborough	Lactating dairy cows	3–5 years old (ca. 600 kg)	Grass silage + concentrates (4 kg DM head ⁻¹ day ⁻¹)
North Wyke	Lactating dairy cows	6 years old (ca. 600 kg)	Concentrate blend, hay, straw, grass silage, maize silage
Pwllpeiran	Lactating dairy cows	6 years old (ca. 600 kg)	Concentrate blend, hay, straw, grass silage, maize silage

month period following application in order to comply with IPCC recommendations (IPCC, 1996).

2.6. Other measurements

2.6.1. Dung and urine composition

Dung and urine sub-samples were taken on the day of application and characterised by measuring pH (in H₂O), dry matter (DM), total N (by Kjeldahl) and total organic carbon content, either using a modified Walkley-Black approach, or analysis by a TOC analyser (uv persulphate oxidation). The readily available N content was also determined, i.e. ammonium N (NH₄⁺-N) and nitrate N (NO₃⁻-N). In addition, two 30 ml sub-samples of urine were taken from each block and preserved by diluting 1:3 with HPLC grade deionised water. The first sample was acidified by adding 1 M H₂SO₄ to reduce the pH to 3 (using a pH meter). To the second sample, 100 µl chloroform was added. Both sub-samples were stored at -20 °C before analysis for urea, hippuric acid, allantoin, uric acid and creatinine, by HPLC (using methods described in Kool et al., 2006).

2.6.2. Soil mineral N and moisture determination

Soil NH₄⁺-N and NO₃⁻-N: Soil samples (0–10 cm) were taken from the dedicated sampling areas of each plot on 10–12 occasions during the 12-month experiment. Fresh soil was passed through a 5 mm sieve before extracting with 2 M KCl and filtering. Filtrates were frozen prior to analysis for NH₄⁺-N and NO₃⁻-N concentrations by colorimetric determination (Singh et al., 2011) using Skalar segmented flow analysers.

Soil moisture content: sub-samples of the sieved soil were weighed (fresh weight) before oven drying at 105 °C overnight, and then reweighed. Soil moisture content was converted to %WFPS using the bulk density of the site (see below) and a particle size density of 2.65 g cm⁻³.

2.6.3. Bulk density

Three representative bulk density measurements were made per site, one per block (walking and sampling a 'W' route across each block), at the start of the experiment, using 100 cm³ bulk density rings, and drying at 105 °C overnight.

2.6.4. Weather data

Daily rainfall and hourly air and soil (0–5 cm) temperature were recorded on site, or daily data used from a nearby weather station (within 1 km) (Table 3).

2.7. Data processing and statistics

The N₂O flux for each chamber was calculated by entering data for the sample vials N₂O concentration, air temperature, closure period and chamber heights into a standard spreadsheet used by all project partners. The mean of the 5 chambers per plot was calculated and used for subsequent calculations of cumulative emissions, using the trapezoidal rule (Cardenas et al., 2010). EFs were calculated by subtracting cumulative N₂O emissions from control plots from treatment plots in the same block. For the urine treatment with DCD the N content in the DCD was taken into account for the calculation of the EF. EFs uniformity of distribution were checked and, if necessary, Box Cox transformation was used on all N₂O data to normalise distribution. Statistical analyses were designed to test:

- the effect of geographical site on N₂O EFs for the different treatments
- the effect of season of application on N₂O EFs for different treatments
- the difference between urine and dung N₂O EFs
- the effect of DCD in reducing N₂O EFs from urine application

Treatment effects and their interactions were evaluated using the F-test in analysis of variance (ANOVA) of each site according to the randomised block design. Multiple comparison of treatment means, if significant, were tested using the Tukey method (Hsu, 1996). When 'treatment × season' interaction was significant then treatments were compared within each season, and seasons were compared with each treatment. In addition, all five sites were combined using REML Meta-analysis in Genstat (VSN International, 2015) where the fixed effects model included main effects and interactions of sites, treatments and seasons (random effects model accounted for the design factors).

Multiple regression analysis (forward selection procedure in Genstat) was used to explore the key soil (% clay, pH, initial % WFPS, average WFPS for first 30 days), environment (average temperature for the first 30 days, average temperature for 365 days after application, total rainfall for the first 30 days, total rainfall for 365 days after application) and urine/dung composition (total urine/dung N content, total urine urea content, total urine/dung ammonium content, uric acid content, hippuric acid content, allantoin content, creatinine content, N application rate) factors that controlled the cumulative N₂O fluxes and N₂O EFs. The main effects of up to (maximum) 10 terms was estimated. No interaction terms were included for selection. In developing a multiple regression model, correlation among the predictor factors (known

Table 3

Weather and soil data for the different urine and dung applications.

Archived data sources: Bell et al. (2017); Cardenas et al. (2017); McGeough et al. (2017); Thorman et al. (2017a, 2017b).

Site	Grazing season period	Measurement period	Whole measurement period		Initial 30 d after application			Day of application		
			Total rainfall (mm)	Average temperature (°C)	Total rainfall (mm)	Average temperature (°C)	Average WFPS (%)	Daily rainfall (mm)	Average daily temperature (°C)	WFPS (%)
Crichton	Early	03/04/12–18/03/13	1325	8.6	60	7.5	51.9	0.1	5.3	41.4
	Mid	27/06/12–10/06/13	1262	8.7	125	14.7	58.3	2.3	12.9	54.8
	Late	08/10/12–25/09/13	1142	9.1	137	7.6	69.8	0.0	8.4	64.6
Drayton	Early	02/05/13–17/04/14	726	10.7	78	10.9	40.1	0.0	10.0	43.6
	Mid	15/08/13–29/07/14	769	9.4	50	15.8	31.0	1.4	20.5	27.5
	Late	17/10/13–14/10/14	787	8.0	123	9.0	45.2	0.0	12.5	42.5
Hillsborough	Early	2/04/12–1/04/13	1191	8.0	77	6.3	84.1	1.4	8.3	88.5
	Mid	25/06/12–24/06/13	1110	8.2	102	14.0	89.3	0.0	12.1	83.1
	Late	17/09/12–16/09/13	1080	8.2	135	8.8	89.9	0.8	10.9	89.7
North Wyke	Early	15/05/12–09/05/13	1405	9.6	143	13.9	57.5	0.6	7.4	58.3
	Mid	03/07/12–11/06/13	1246	9.4	103	13.6	62.0	7.6	15.4	62.4
	Late	26/09/12–10/09/13	1288	9.5	160	9.1	63.1	11.1	12.4	71.6
Pwllpeiran	Early	11/04/13–27/03/14	1878	10.4	96	8.4	44.8	5.7	7.8	44.1
	Mid	04/07/13–16/06/14	1844	10.8	61	17.2	36.7	0.0	14.4	44.1
	Late	12/09/13–27/08/14	1841	10.6	98	13.6	44.8	13.6	14.4	42.2

as multicollinearity) can affect model equation stability. For this modelling exercise, we used the statistical package Genstat (Genstat 18th Ed.; VSN International, 2015), which has the built-in facility to check for any multicollinearity issues (any such problem can be dealt with by using Genstat Procedure “Ridge” regression which incorporates Principal Component (PCA) regression).

3. Results

3.1. Urine and dung composition

The N content of the urine used in the 15 experiments (Table 4a) were typical for cattle urine (Dijkstra et al., 2013; Selbie et al., 2015; Gardiner et al., 2016), ranging from 6.8 to 11.4 g l⁻¹ (average 9.11 g l⁻¹ ± 0.35). In most cases urea-N represented between 60 and 100% of the total N content. However, for the three experiments at Hillsborough, the low urea-N content of the urine was linked to a high urine ammonium-N content (Table 4a), indicating hydrolysis of urea prior to application to the soil. Since urea hydrolysis is such a rapid process once urine has been deposited on the soil, we do not consider the N₂O emissions from the three Hillsborough experiments to have been directly affected by this.

Concentrations of the purine derivatives in the urine varied markedly between the different seasons of collection for the different experiments at each site, and between sites (Table 5).

This reflects differences in the diets that cattle were fed prior to collection of the urine on each occasion (see Table 2 for a summary of the diets), and differences between cattle groups at each collection site. However, concentrations are typical of those reported in the literature (Dijkstra et al., 2013; Selbie et al., 2015; Gardiner et al., 2016). The measured N contained in the purine derivatives represented from 3 to 28% of the total N content of the urine (average 12.5% ± 0.02).

The total N content of the dung ranged from 3.4 to 48.0 g kg⁻¹ (DM), whilst the DM content ranged from 10.6–36.2% (Table 4b). The total N loadings in the urine and dung treatments were typical for cattle, 338–568 kg ha⁻¹ (average 455 ± 17.6) and 625–1020 kg ha⁻¹ (average 835 ± 31.9), respectively. These values are within reported ranges (Selbie et al., 2015).

3.2. Weather

Annual rainfall was greater than the 30-year mean in two (of the three) Crichton experiments, and all three experiments at Drayton, Hillsborough, North Wyke and Pwllpeiran. Average annual air

Table 4b

Average dung composition, and N and C loading rates for each experiment. DM = dry matter.

Archived data sources: Bell et al. (2017); Cardenas et al. (2017); McGeough et al. (2017); Thorman et al. (2017a, 2017b).

	Grazing season period	Total N loading (kg ha ⁻¹)	pH	DM (%)	Total N (g kg ⁻¹ DM)	NH ₄ ⁺ -N (mg kg ⁻¹ DM)	NO ₃ ⁻ -N (mg kg ⁻¹ DM)
Crichton	Early	1020	–	12.9	5.10	410	0.0
	Mid	680	–	11.5	3.40	260	0.0
	Late	720	–	10.6	3.60	230	0.0
Drayton	Early	840	7.6	18.9	22.2	5020	24.5
	Mid	736	7.6	36.2	10.2	3680	6.9
	Late	802	7.8	27.0	14.8	4443	0.0
Hillsborough	Early	980	6.9	14.5	4.90	500	0.0
	Mid	976	7.3	14.3	4.90	669	0.0
	Late	1008	7.7	14.3	5.00	683	0.0
North Wyke	Early	911	7.0	14.5	31.4	3035	0.1
	Mid	625	7.4	21.1	48.0	4310	21.6
	Late	771	7.5	20.5	18.8	2940	20.8
Pwllpeiran	Early	769	7.5	24.5	15.7	5095	25.2
	Mid	823	7.3	23.3	17.7	6497	7.9
	Late	866	7.5	24.1	18.0	5833	3.5

temperature was similar to the 30-year mean at Crichton and Pwllpeiran, cooler at Hillsborough and North Wyke, and warmer at Drayton. However, it is more likely that the weather conditions immediately before urine and dung application, and within the first three months after application would have the most influence on N₂O production and emission (see Table 3).

3.3. Nitrous oxide emissions

3.3.1. Controls

Background (control) cumulative N₂O emissions ranged from –0.03–1.26 kg N₂O-N ha⁻¹ for all sites and all experiments, with an average from the data in Table 6 of 0.49 kg N₂O-N ha⁻¹ (±0.10). From the meta-analysis, we find that across all seasons, the N₂O emissions from the controls were significantly greater from the Crichton, North Wyke and Pwllpeiran sites compared to the Drayton site (*p* < 0.05). Within an individual site, emissions from controls also varied between seasons of application, particularly at the North Wyke site. There was no statistically significant relationship between the urine N₂O EF and the cumulative annual N₂O emission from the control plots (*p* > 0.05). Across all sites, N₂O emissions from the control plots at the early grazing

Table 4a

Average urine composition, and N and C loading rates for each experiment.

For urine + DCD treatments, an additional 6.5 kg N ha⁻¹ was supplied in the inhibitor. DM = dry matter.

Archived data sources: Bell et al. (2017); Cardenas et al. (2017); McGeough et al. (2017); Thorman et al. (2017a, 2017b).

Site	Grazing season period	Urine							Artificial urine						
		Total N loading (kg ha ⁻¹)	pH	DM (%)	Total N (g l ⁻¹)	Urea-N (mg l ⁻¹)	NH ₄ ⁺ -N (mg l ⁻¹)	NO ₃ ⁻ -N (mg l ⁻¹)	Total N loading (kg ha ⁻¹)	pH	DM (%)	Total N (g l ⁻¹)	Urea-N (mg l ⁻¹)	NH ₄ ⁺ -N (mg l ⁻¹)	NO ₃ ⁻ -N (mg l ⁻¹)
Crichton	Early	480	–	4.9	9.60	6332	120	–	180	–	1.4	3.60	1318	–	–
	Mid	420	–	4.6	8.40	8127	240	–	425	–	3.5	8.50	9264	–	–
	Late	435	–	4.9	8.70	6231	100	–	425	–	3.5	8.50	9264	–	–
Drayton	Early	540	7.5	5.5	10.80	10,780	825	0.5	501	7.1	4.6	10.01	9820	25	0.5
	Mid	454	9.0	4.5	9.07	8540	4870	1.5	495	7.3	3.9	9.91	8340	39	0.0
	Late	471	8.1	5.2	9.43	8480	315	0.0	495	7.1	4.7	9.90	10,040	25	0.0
Hillsborough	Early	432	9.0	5.5	8.64	2900	6917	0.0	510	7.7	4.6	10.20	7335	100	115.0
	Mid	338	8.9	5.3	6.75	375	5862	27.0	502	7.6	5.0	10.04	8035	55	126.0
	Late	354	9.0	4.2	7.07	767	6216	41.0	504	8.2	4.5	10.08	8048	88	163.0
North Wyke	Early	405	8.3	5.3	8.10	6521	554	0.0	440	8.2	4.3	8.80	7079	18	0.1
	Mid	429	7.3	4.8	8.57	6284	1230	1.0	481	7.5	4.2	9.61	6833	<50	0.4
	Late	435	9.2	4.5	8.70	7382	2020	2.5	423	7.4	3.4	8.45	7774	<50	0.8
Pwllpeiran	Early	565	9.3	5.6	11.30	10,100	2743	0.3	495	9.3	4.4	9.91	9620	315	0.3
	Mid	568	7.8	5.5	11.37	6840	822	0.3	498	7.4	3.9	9.96	7840	25	0.2
	Late	505	7.8	4.7	10.10	8820	115	0.3	508	7.5	4.1	10.15	10,040	25	0.0

Table 5

Concentrations (g l^{-1}) of purine derivatives in cattle urine used in the experiments. *Detection limit of the analytical approach. To convert from $\text{mg molecule l}^{-1}$ to mg N l^{-1} , multiply hippuric acid by 0.078138, allantoin by 0.354161, uric acid by 0.333115, and creatinine by 0.371287. Archived data sources: Bell et al. (2017); Cardenas et al. (2017); McGeough et al. (2017); Thorman et al. (2017a, 2017b).

Site	Grazing season period	Hippuric acid	Allantoin	Uric acid	Creatinine
Crichton	Early	9.17	2.42	0.30	0.68
	Mid	1.57	1.70	0.45	1.29
	Late	7.69	3.89	0.41	0.77
Drayton	Early	<0.50*	2.83	0.48	0.58
	Mid	<0.50*	<0.40*	0.55	0.62
	Late	8.02	3.51	0.54	0.68
Hillsborough	Early	<0.50*	0.74	0.15	0.40
	Mid	<0.50*	<0.40*	0.06	<0.10*
	Late	<0.50*	<0.40*	0.12	<0.10*
North Wyke	Early	3.92	1.91	0.37	0.76
	Mid	<0.50*	<0.40*	0.40	0.52
	Late	4.86	<0.40*	0.35	0.52
Pwllpeiran	Early	5.13	0.84	0.36	0.81
	Mid	<0.50*	<0.40*	0.31	0.25
	Late	8.92	3.67	0.03	0.73

application timing were significantly greater than from the late-grazing application ($p < 0.05$). Regression modelling indicated that the key factors controlling the magnitude of the annual N_2O fluxes from control plots were soil organic carbon content, clay content, bulk density, WFPS during the first 30d after application, and average annual temperature, with these factors accounting for ca. 56% of the variance in emissions. The resulting full regression equation was: Cumulative N_2O flux

$$(\text{kg N ha}^{-1}) = 3.981 - 0.0846 \text{ SOC} - 0.02220 \text{ initial WFPS} + 0.01052 \times 30\text{d WFPS} - 1.683 \text{ Bulk density} - 0.01807 \text{ Clay content} - 0.0408 \times 365\text{d average temperature}.$$

3.3.2. Urine

Examples of daily N_2O fluxes are shown in Fig. 3 for the late-season urine, dung and control treatments at the Drayton site. These data show two distinct peaks in N_2O fluxes, something observed in several of the experiments (e.g. Cardenas et al., 2016), suggesting the peaks in emission are associated either with different processes (e.g. denitrification of soil NO_3 during the first peak as a result of the carbon addition in the urine, and nitrification of the urine NH_4 source during the second peak), or different pools of N being the substrate for denitrification (e.g. the first peak associated with the urine-derived NH_4 , and the second peak associated with other more recalcitrant pools, e.g. N contained in purine derivatives). Further research using labelled urine N compounds would help reveal the underpinning processes and/or N sources responsible for the two peaks in emission.

The mean urine N_2O EF was 0.69% (± 0.20), ranging from 0.05–2.96 (Table 6). Across all seasons of application, the meta-analysis showed that the N_2O EF was significantly greater from the North Wyke site than other sites ($p < 0.05$) (Fig. 4). Whilst across all sites, the N_2O EF was significantly greater following an early-grazing application ($p < 0.05$) (Fig. 5). DCD reduced the N_2O EF from urine in 13 of the 15 experiments, although this reduction was only significant in 5 of these experiments (Table 6). The average N_2O EF for the urine + DCD treatment was 0.37% (± 0.09) (Table 6). So, the use of DCD resulted in an average reduction in the N_2O EF of 46%, although the range in efficacy was wide, i.e. from an increase in the N_2O EF of 32% (mid-season application at Hillsborough) to a reduction of 75% (at the same site from the early-season application).

Table 6

Average cumulative N_2O emissions and N_2O EFs from the urine and dung treatments at each experimental site for each application. (Values in italics are standard errors of the mean). Archived data sources: Bell et al. (2017); Cardenas et al. (2017); McGeough et al. (2017); Thorman et al. (2017a, 2017b). Within each site/timing experiment (rows), average total N_2O emissions or N_2O EFs between excretal N sources with different letters are significantly different ($p < 0.05$, $N = 3$).

Site	Grazing season period	Cumulative emissions of N ₂ O (kg N ₂ O ha ^{−1})					N ₂ O EF (% of applied N)			
		Control	Urine	Urine +DCD	Artificial urine	Dung	Urine	Urine +DCD	Artificial urine	Dung
Crichton	Early	0.96a	1.92a	1.25a	0.93a	2.15a	0.20a	0.06a	−0.02a	0.12a
		0.23	0.20	0.09	0.09	0.51	0.06	0.03	0.13	0.03
	Mid	0.61a	5.18b	5.07b	5.29b	2.00a	1.09b	1.05b	1.10b	0.20a
		0.17	0.82	1.03	0.30	0.09	0.18	0.28	0.06	0.03
	Late	0.79a	2.21a	1.79a	1.49a	1.55a	0.33a	0.23a	0.16a	0.11a
		0.32	0.66	0.24	0.60	0.12	0.15	0.12	0.18	0.03
Drayton	Early	0.18a	2.02a	1.35a	1.86a	0.85a	0.34a	0.21a	0.34a	0.08a
		0.06	0.11	0.14	0.05	0.17	0.03	0.02	0.01	0.02
	Mid	0.03a	0.86a	0.74a	0.82a	0.95a	0.18a	0.15a	0.16a	0.12a
		0.09	0.08	0.07	0.08	0.05	0.00	0.00	0.01	0.01
	Late	−0.03a	7.68d	4.73bc	6.47 cd	2.56b	1.64c	1.00b	1.31bc	0.32a
		0.05	1.79	1.35	0.42	0.35	0.37	0.28	0.08	0.04
Hillsborough	Early	0.36a	4.78a	1.46a	10.87b	1.98a	1.02a	0.25a	2.06b	0.17a
		0.10	1.11	0.42	4.84	0.20	0.26	0.12	0.96	0.03
	Mid	0.23a	1.20a	1.52a	1.96a	1.73a	0.29a	0.38a	0.34a	0.15a
		0.04	0.15	0.44	0.61	0.45	0.05	0.14	0.13	0.04
	Late	0.15a	0.31a	0.20a	0.67a	0.51a	0.05a	0.01a	0.10a	0.04a
		0.05	0.07	0.04	0.11	0.15	0.03	0.00	0.03	0.01
North Wyke	Early	1.26a	13.26d	5.54b	11.06c	2.50a	2.96c	1.09b	2.23c	0.14a
		0.13	0.50	0.69	0.43	0.43	0.14	0.20	0.12	0.06
	Mid	0.80a	3.19b	2.93b	4.16b	3.24b	0.56b	0.49ab	0.70b	0.39a
		0.07	0.51	0.50	0.89	0.52	0.11	0.10	0.18	0.08
	Late	0.03a	0.52a	0.59a	0.34a	0.82a	0.11a	0.12a	0.07a	0.10a
		0.13	0.29	0.11	0.23	0.17	0.04	0.02	0.02	0.01
Pwllpeiran	Early	0.49a	3.45c	1.59ab	3.15c	2.17bc	0.52b	0.19a	0.54b	0.22a
		0.18	0.34	0.31	0.23	0.31	0.07	0.07	0.03	0.05
	Mid	0.42a	2.11b	0.94ab	1.81b	2.13b	0.30b	0.09a	0.28b	0.21ab
		0.04	0.13	0.03	0.15	0.33	0.03	0.01	0.03	0.04
	Late	0.52a	4.14b	1.78a	3.52b	5.08c	0.72c	0.25a	0.59bc	0.53b
		0.07	0.39	0.13	0.36	0.81	0.09	0.02	0.08	0.09
Average of all sites and seasons							0.69	0.37	0.66	0.19
							0.20	0.09	0.18	0.03

Table 7
Significance F-test probabilities for cumulative N₂O emission and N₂O EF, by timing of application, site, and timing of application × site interactions, from randomised block design ANOVA for each experiment.

		Treatments	Application time	Interaction
Crichton	Total N ₂ O	<0.001	<0.001	<0.001
	EF	0.012	<0.001	0.019
Drayton	Total N ₂ O	<0.001	<0.001	<0.001
	EF	<0.001	<0.001	0.002
Hillsborough	Total N ₂ O	0.006	<0.001	0.014
	EF	0.035	0.002	0.042
North Wyke	Total N ₂ O	<0.001	<0.001	<0.001
	EF	<0.001	<0.001	<0.001
Pwllpeiran	Total N ₂ O	<0.001	<0.001	<0.001
	EF	<0.001	<0.001	0.093

3.3.3. Artificial urine

The mean artificial urine N₂O EF was similar to that of the real urine, 0.66% (± 0.18) (Table 6), and there was a good relationship between the N₂O EFs for real and artificial urine ($r^2 = 0.77$). Across all seasons, the meta-analysis showed that the N₂O EF from the artificial urine was significantly greater at North Wyke and Hillsborough ($p < 0.05$) than the other sites (Fig. 4). Across all sites, the greatest N₂O EF occurred following the early-grazing application ($p < 0.05$) (Fig. 5).

3.3.4. Dung

The mean N₂O EF for dung (from the meta-analysis) was 0.19% (± 0.03), with a range of 0.04–0.53 (Table 6), which was significantly lower than for urine ($p < 0.05$). The meta-analysis showed there was no effect of site or season of application on the N₂O EF from dung ($p > 0.05$) (Figs. 4 and 5).

3.4. Factors affecting N₂O fluxes from urine and dung

It is clear that there were significant ($p < 0.05$) effects of excretal N source and season of application at each site, as well as ‘treatment’ × ‘season’ interactions (Table 7).

3.4.1. Urine

Multiple regression analysis showed that the factors that best explained cumulative N₂O emissions from urine application mainly included urine composition and soil pH. The factors explaining 91.1% of the variance in cumulative N₂O emissions from urine patches are shown via this equation: Cumulative N₂O flux (kg N ha^{-1}) = $-61.94 + 38.50$ urine creatinine content -0.0042 urine urea N content $+0.003310$ urine ammonium N content $+0.002801$ urine total nitrogen content $+4.115$ soil pH -1.036 urine hippuric acid content $+4.340$ urine pH -8.06 urine uric acid content. >75% of the variance in total

N₂O flux was explained by the urine total N, urea-N, ammonium-N, uric acid and creatinine content.

The full equation of factors explaining 91.1% of the urine N₂O EF was; $\text{EF\%} = -15.9 + 8.776$ urine creatinine content -0.0009595 urine urea N content -0.0007965 urine ammonium N content $+1.014$ soil pH $+0.0005941$ urine total nitrogen content -0.2563 urine hippuric acid content $+1.116$ urine pH -2.059 urine uric acid content. >75% of the variance in N₂O EF was explained by the urine total N, urea-N, ammonium-N, uric acid and creatinine content.

3.4.2. Dung

In contrast to urine, multiple regression showed that the factors that best explained cumulative N₂O emissions from dung application included environmental and soil factors (as well as dung factors). The full equation, explaining 68.3% of the variance in cumulative N₂O emissions from dung in this study was; Cumulative N₂O flux (kg N ha^{-1}) = $4.15 - 0.0579$ initial %WFPS -0.308 365d average temperature -0.805 soil pH -0.0408 dung nitrate N content -0.00082 total nitrogen applied $+1.053$ soil organic carbon -10.50 soil dry bulk density $+1.927$ dung pH.

The full equation of factors explaining 66.5% of the dung N₂O EF was; $\text{EF\%} = -0.295 + 0.0001187$ dung ammonium N content $+0.01784$ 30d %WFPS -0.01473 dung nitrate N content -0.002143 total nitrogen applied -0.02343 30d average temperature $+0.1159$ soil organic carbon $+0.1747$ dung total nitrogen content $+0.0452$ 365d average temperature.

4. Discussion

Urine N₂O EFs were significantly greater (average 0.69%) than the dung N₂O EFs (average 0.19%), signifying the importance of the Nr content as a substrate for the soil processes, nitrification and denitrification, responsible for N₂O production. Our urine and dung N₂O EFs are similar

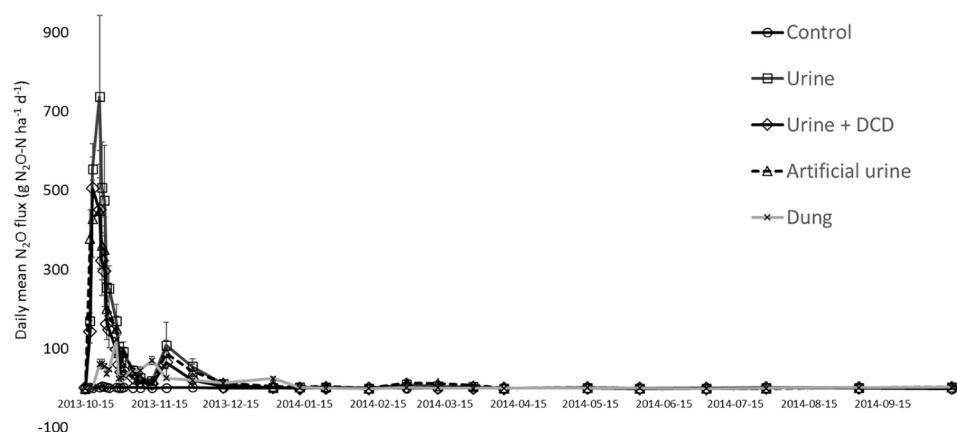


Fig. 3. Daily mean N₂O fluxes following urine and dung treatments at Drayton after a late-season application. ($N = 3$, vertical bars are standard error of the mean).

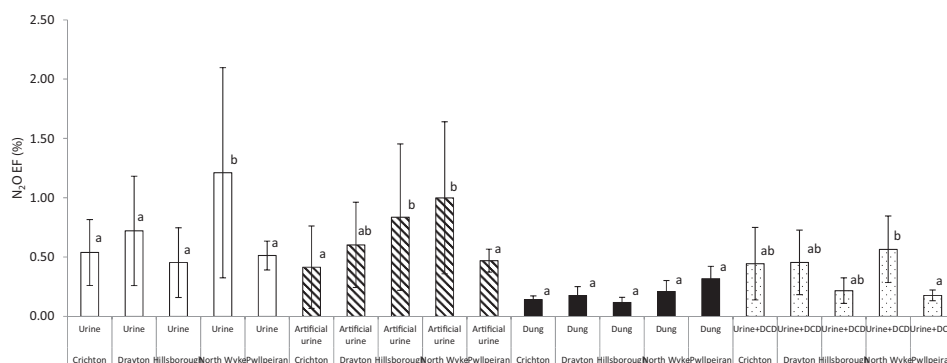


Fig. 4. Average N₂O EF (across three seasons of application) for each site, for urine and dung treatments. Within each urine/dung treatment, average N₂O EFs (from meta-analysis) between sites with different letters are significantly different ($N = 3$, vertical lines are standard error of the mean).

to some of those measured by New Zealand researchers, summarised by Kelliher et al. (2014). In New Zealand, urine N₂O EFs are categorised by livestock species and farming system (lowland, hill country low and high slope), and our results are more similar to the N₂O EFs for the hill-country low slope dairy cattle urine (average of 0.84%) and dung (average of 0.20%). By contrast, Krol et al. (2016) reported larger average urine and dung N₂O EFs for nine experiments conducted in Ireland of 1.18% (urine) and 0.39% (dung); EFs approximately double the values we have measured. In this series of experiments, Krol et al. (2016) applied urine at a higher N loading rate (average of 720 kg N ha⁻¹) than in our study (average of 455 kg N ha⁻¹). However, the greater N₂O EF from the dung in the Irish study (0.39%) was despite using a lower N loading rate (average of 459 kg N ha⁻¹) than in our study (835 kg N ha⁻¹), suggesting that N loading was not the only factor resulting in the greater urine N₂O EFs in these Irish experiments. Soil and environmental factors appeared to have been more conducive to N₂O production and emission in this Irish study.

In our study, DCD reduced the urine N₂O EFs by an average of 46%, although there was considerable variability in its efficacy to reduce N₂O emissions (between sites and between seasons). In a related study, McGeough et al. (2016) took soil from these five UK grassland sites, and an additional four arable sites, and demonstrated that the efficacy of DCD to inhibit nitrification was controlled by the interaction between temperature, soil clay content and soil organic matter. Moreover, this study concluded that DCD was more effective in arable soils than in these grassland soils (McGeough et al., 2016). The average DCD N₂O mitigation efficacy we measured (46%), and the range of efficacy that we measured are similar to other studies. For example, Selbie et al. (2014) showed that DCD increased the urine N₂O EF by an average

of +4% (a small increase) for urine applied at a loading rate of 500 kg N ha⁻¹, but resulted in a 30% reduction for urine applied at 1000 kg N ha⁻¹ (in New Zealand). Misselbrook et al. (2014) reported a greater efficacy of DCD to reduce the urine N₂O EF, by 70% on a sandy clay loam in SW England. Recently, Minet et al. (2018) showed DCD, applied at 10 kg ha⁻¹, could reduce the urine N₂O EF by 34% (from 0.80% to 0.52%), but that DCD applied at 30 kg ha⁻¹ reduced the urine N₂O EF further, by 64%. Note: efficacy of DCD is often reported for cumulative emissions, with reported values being much higher than the efficacy of reducing the EF itself (e.g. Selbie et al., 2014). However, the efficacy of DCD to reduce N₂O EFs is needed if national inventories are to be modified accordingly.

We found evidence of the effect of timing on N₂O EFs, with larger EFs occurring following early-season urine application/deposition (Fig. 5). Krol et al. (2016) also explored the effect of season of urine application on N₂O EFs from Irish grasslands, and showed that EFs varied seasonally, with the highest EFs in the autumn, and that emission were also dependent on soil type. Indeed, relationships between the magnitude of N₂O EFs with “generic” season of deposition should be interpreted with caution, as soil and environmental conditions can vary markedly within a season. Hence, the importance of using statistical regression modelling to explore the key controls. Whilst there were insufficient data from our 15 experiments to be able to explore the relationships between cumulative N₂O emissions, N₂O EFs and climate/soil with certainty, the limited regression analysis showed that N₂O emissions associated with urine were more related to urine composition than environmental and soil factors, whilst for dung which has a relatively low inorganic N content, N₂O emissions were also controlled by soil and environmental factors. Krol et al. (2016) also used regression modelling to show the

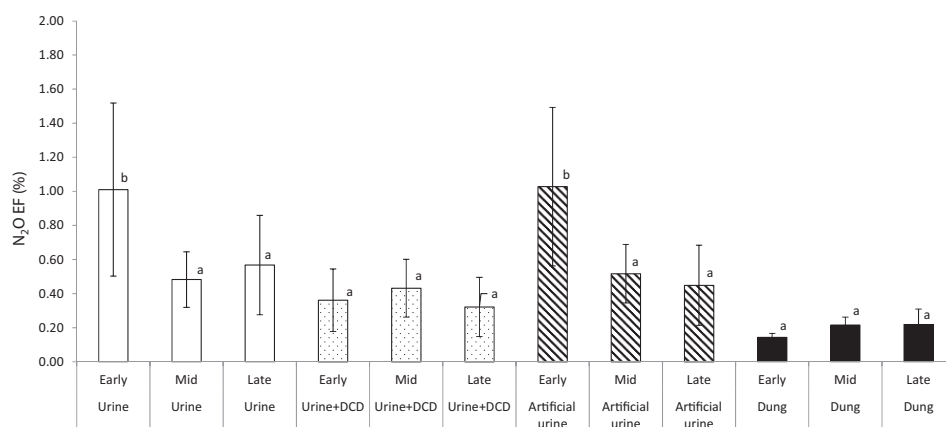


Fig. 5. Effect of urine/dung treatment application timing (across all sites) on average N₂O EF. Within each treatment, average N₂O EFs (from meta-analysis) between timings of application with different letters are significantly different ($N = 3$, vertical lines are standard error of the mean).

importance of rainfall and temperature before, and soil moisture deficit after, application of excretal deposition, on N_2O emissions from nine experiments on Irish grasslands. We recognise the limitations of conducting regression analysis on such small data sets. However, there is potential to generate a much larger data set by combining data from studies where soils and climate are similar, and where similar protocols were followed, e.g. Krol et al. (2016), Minet et al. (2018), and data from some New Zealand experiments, to explore the controls of N_2O emissions from urine and dung deposition, and generate improved EFs. Importantly, our unique dataset of daily N_2O fluxes, cumulative emissions and emission factors, as well as soil mineral N and moisture data with weather, soil and site information have all been archived for future use by researchers (Bell et al., 2017; Cardenas et al., 2017; McGeough et al., 2017; Thorman et al., 2017a; Thorman et al., 2017b), and to allow integration with future datasets that become available.

To calculate a provisional excretal N_2O EF, based on the data presented in this study, we assume a 60:40 split between the total N excreted in urine and dung (Webb and Misselbrook, 2004). We estimate a combined excretal N_2O EF, based on our mean urine and dung N_2O EFs data of 0.49%. These UK data have now been combined with the very few additional IPCC compliant UK experimental datasets (see Misselbrook et al., 2014) to generate a new country specific N_2O EF of 0.44%. This is <25% of the IPCC (2006) default EF for cattle grazing excreta (EF_3), and ca. 50% of the default EF for sheep grazing excreta. If we substitute this new pasture, range and paddock EF for both cattle and sheep into the IPCC, 2006 methodology for calculating the UK inventory, we estimate a reduction of 11.6 kt N_2O (18% less N_2O for UK agriculture for 2015) and for total UK agricultural GHG emissions, a reduction of 3.4 Mt. CO_2e , or 7% for UK agriculture for 2015. This new EF is used in back-casting to 1990, and so has no bearing on meeting the UK's ambitious greenhouse gas mitigation target. However, a reduced GHG emission from agriculture means that a greater proportion of the emission can be "offset" by carbon sequestration, and suggests that e.g. land sparing strategies may be more realistic (Lamb et al., 2016). The lower country specific pasture, range and paddock EF_3 also has implications for calculating carbon footprints of ruminant livestock products in the UK.

Clearly, this study focussed on cattle urine and dung where applications were made to lowland mineral soils, and where urine and dung were collected from cattle fed "lowland" diets. So, questions arise about a) extrapolating the N_2O EF data to sheep; indeed the IPCC default sheep urine N_2O EF (1%) is greater than the new combined cattle excreta N_2O EF from our study, and b) extrapolating the new N_2O EF data to beef and sheep grazing in the uplands, on much more organic and potentially acidic soils, and where weather and soil conditions as well as urine/dung composition may be very different.

5. Conclusions

This was the first co-ordinated study in the UK to generate data to develop a country specific grazing excreta N_2O EF for cattle. Results confirmed that urine is the greatest source of N_2O compared to dung, and that the nitrification inhibitor, DCD, offers the potential to reduce N_2O emissions from urine patches, although its efficacy across the sites and seasons was variable. Understanding what controls this variability, and the development of cost effective delivery mechanisms need to be addressed if this technology is to be adopted. Importantly, the results of this study provide evidence that for the UK soil and climatic conditions, the N_2O EF for grazing excreta for cattle is significantly lower (0.49%) than the IPCC default (2%) with implications for both government and the ruminant livestock industries as they seek to meet challenging greenhouse gas mitigation targets and greenhouse gas emission roadmaps, respectively. Further questions arise in terms of the validity of extrapolating these data from cattle to sheep grazing, and from mineral to organic soils.

Data statement

Our unique datasets of daily N_2O fluxes, cumulative emissions and emission factors, as well as soil mineral N and moisture data with weather, soil and site information from all 15 experiments have been archived for future use by researchers (see: Bell et al., 2017; Cardenas et al., 2017; McGeough et al., 2017; Thorman et al., 2017a; Thorman et al., 2017b), and to allow integration with future datasets that become available.

Acknowledgements

The authors are grateful to the UK Department for Environment, Food and Rural Affairs (Defra), the Department of Agriculture and Rural Development (now the Department of Agriculture, Environment and Rural Affairs) in Northern Ireland, and the Scottish Government and the Welsh Government for financial support via the *InveN2Ory* project (AC0116). The work by Rothamsted Research was additionally supported by the Biotechnology and Biological Sciences Research Council (BBS/E/C/00010320). We would also like to thank Jon Moorby (IBERS, Wales, UK), Reading University, SRUC (Scotland, UK) and Conrad Ferris (AFBI, Northern Ireland, UK), for provision of cattle urine and dung.

References

- Balvert, S., Luo, J., Schipper, L., 2017. Do glucosinolate hydrolysis products reduce nitrous oxide emissions from urine affected soil? *Sci. Total Environ.* 603–604, 370–380.
- Barneze, A.S., Minet, E.P., Cerri, C.C., Misselbrook, T., 2015. The effect of nitrification inhibitors on nitrous oxide emissions from cattle urine depositions to grassland under summer conditions in the UK. *Chemosphere* 119, 122–129.
- Bell, M., Rees, R., Cloy, J., Topp, K., Bagnall, A., Chadwick, D., 2015. Nitrous oxide emissions from cattle excreta applied to a Scottish grassland: effects of soil and climatic conditions and a nitrification inhibitor. *Sci. Total Environ.* 508, 343–353.
- Bell, M.J., Rees, R.M., Cloy, J.M., Topp, C.F.E., Bagnall, A., Chadwick, D.R., 2017. Agricultural greenhouse gas inventory research platform - *InveN2Ory*. Dung and urine experimental site in Dumfries, 2012. Version: 1. [dataset] Freshwater Biological Association [publisher]. <https://doi.org/10.17865/ghgno578>.
- Broderick, G.A., 2003. Effects of varying dietary protein and energy levels on the production of lactating dairy cows. *J. Dairy Sci.* 86, 1370–1381.
- Burchill, W., Lanigan, G.J., Forrester, P.J., Misselbrook, T., Richards, K.G., 2017. Ammonia emissions from urine patches amended with N stabilized fertilizer formulations. *Nutr. Cycl. Agroecosyst.* 108, 163–175.
- Cardenas, L.M., Thorman, R., Ashlee, N., Butler, M., Chadwick, D.R., Chambers, B., Cuttle, S., Donovan, N., Kingston, H., Lane, S., Scholefield, D., 2010. Quantifying annual N_2O emission fluxes from grazed grassland under a range of inorganic fertiliser nitrogen inputs. *Agric. Ecosyst. Environ.* 136, 218–226.
- Cardenas, L.M., Misselbrook, T.M., Hodgson, C., Donovan, N., Gilhespy, S., Smith, K.A., Dhanoa, M.S., Chadwick, D., 2016. Effect of the application of cattle urine with or without the nitrification inhibitor DCD, and dung on greenhouse gas emissions from a UK grassland soil. *Agr. Ecosyst. Environ.* 235, 229–241.
- Cardenas, L.M., Misselbrook, T.H., Donovan, N., 2017. Agricultural Greenhouse Gas Inventory Research Platform - *InveN2Ory*. Dung and urine experimental site in Devon, 2012. Version: 1. [dataset] Freshwater Biological Association [publisher]. <https://doi.org/10.17865/ghgno562>.
- Chadwick, D.R., Rees, R.M., Williams, J., Smith, P., Skiba, U.M., Hiscock, K., Manning, A.J., Watson, C., Smith, K.A., Anthony, S.G., Moorby, J., Mottram, T., 2011. Improving the national inventory of agricultural nitrous oxide emissions from the UK (*InveN2Ory*). Non- CO_2 Greenhouse Gases (NCGG-6) Science, Policy and Integration (Conference Proceedings). 2011. Amsterdam.
- Chadwick, D.R., Cardenas, L., Misselbrook, T.H., Smith, K.A., Rees, R.M., Watson, C.J., McGeough, K.L., Williams, J.R., Cloy, J.M., Thorman, R.E., Dhanoa, M.S., 2014. Optimizing chamber methods for measuring nitrous oxide emissions from plot-based agricultural experiments. *Eur. J. Soil Sci.* 65, 295–307.
- Clough, T.J., Ledgard, S.F., Sprosen, M.S., Kear, M.J., 1998. Fate of N-15 labelled urine on four soil types. *Plant Soil* 199, 195–203.
- Clough, T.J., Ray, J.L., Buckthought, L.E., Calder, J., Baird, D., O'callaghan, M., Sherlock, R.R., Condron, L.M., 2009. The mitigation potential of hippuric acid on N_2O emissions from urine patches: an in situ determination of its effect. *Soil Biol. Biochem.* 41, 2222–2229.
- de Klein, C.A.M., Ledgard, S.F., 2001. An analysis of environmental and economic implications of nil and restricted grazing systems designed to reduce nitrate leaching from New Zealand dairy farms. I. Nitrogen losses. *N. Z. J. Agric. Res.* 44, 201–215.
- de Klein, C.A., Luo, J., Woodward, K.B., Styles, T., Wise, B., Lindsey, S., Cox, N., 2014. The effect of nitrogen concentration in synthetic cattle urine on nitrous oxide emissions. *Agric. Ecosyst. Environ.* 188, 85–92.
- Di, H.J., Cameron, K.C., 2007. Nitrate leaching losses and pasture yields as affected by different rates of animal urine nitrogen returns and application of a nitrification inhibitor - a lysimeter study. *Nutr. Cycl. Agroecosyst.* 79, 281–290.

- Di, H.J., Cameron, K.C., 2008. Sources of nitrous oxide from ^{15}N -labelled animal urine and urea fertiliser with and without a nitrification inhibitor, dicyandiamide (DCD). *Aust. J. Soil Res.* 46, 76–82.
- Di, H.J., Cameron, K.C., 2012. How does the application of different nitrification inhibitors affect nitrous oxide emissions and nitrate leaching from cow urine in grazed pastures? *Soil Use Manag.* 28, 54–61.
- Dijkstra, J., Oenema, O., Van Groenigen, J.W., Spek, J.W., Van Vuuren, A.M., Bannink, A., 2013. Diet effects on urine composition of cattle and N_2O emissions. *Animal* 7, 292–302.
- Gardiner, C.A., Clough, T.J., Cameron, K.C., Di, H.J., Edwards, G.R., de Klein, C.A.M., 2016. Potential for forage diet manipulation in New Zealand pasture ecosystems to mitigate ruminant urine derived N_2O emissions: a review. *N. Z. J. Agric. Res.* 59, 301–317.
- Hatch, D., Trindad, H., Cardenas, L., Carneiro, J., Hawkins, J., Scholefield, D., Chadwick, D., 2005. Laboratory study of the effects of two nitrification inhibitors on greenhouse gas emissions from a slurry-treated arable soil: impact of diurnal temperature cycle. *Biol. Fertil. Soils* 41, 225–232.
- Hsu, J.C., 1996. *Multiple Comparisons Theory and Methods*. Chapman & Hall, London.
- IPCC, 1996. Intergovernmental panel on climate change, 1996. Revised 1996 IPCC Guidelines for National Greenhouse Gas Inventories. IPCC/OECD/IEA, United Kingdom.
- IPCC, 2000. Intergovernmental Panel on Climate Change, 2000. Good practice guidance and uncertainty management in national greenhouse gas inventories. In: Penman, J., Kruger, D., Galbally, I., Hiraishi, T., Nyenzi, B., Emmanuel, S., Buendia, L., Hoppaus, R., Martinsen, T., Meijer, J., Miwa, K., Tanabe, K. (Eds.), Intergovernmental Panel on Climate Change (IPCC). IPCC/OECD/IEA/IGES, Hayama, Japan.
- IPCC, 2006. Chapter 11: N_2O Emissions from Managed Soils, and CO_2 Emissions from Lime and Urea Application. In: De Klein, C., Novoa, R.S.A., Ogle, S., Smith, K.A., Rochette, P., Wirth, T.C., McConkey, B.G., Mosier, A., Rypdal, K. (Eds.), Intergovernmental Panel on Climate Change, 2006. Guidelines for National Greenhouse Gas Inventories Agriculture, Forestry and Other Land Use Intergovernmental Panel on Climate Change http://www.ipcc-nggip.iges.or.jp/public/2006gl/pdf/4_Volume4/V4_11_Ch11_N2O&CO2.pdf, Accessed date: 2 December 2018.
- Kelliher, F.M., Cox, N., van der Weerden, T.J., de Klein, C.A.M., Luo, J., Cameron, K.C., Di, H.J., Giltrap, D., Rys, G., 2014. Statistical analysis of nitrous oxide emission factors from pastoral agricultural field trials conducted in New Zealand. *Environ. Pollut.* 186, 83–86.
- Kool, D.M., Hoffland, E., Abrahamse, S.P.A., van Groenigen, J.W., 2006. What artificial urine composition is adequate for simulating soil N_2O fluxes and mineral N dynamics? *Soil Biol. Biochem.* 38, 1757–1763.
- Krol, D.J., Carolan, R., Minet, E., McGeough, K.L., Watson, C.J., Forrester, P.J., Lanigan, G.J., Richards, K.G., 2016. Improving and disaggregating N_2O emission factors for ruminant excreta on temperate pasture soils. *Sci. Total Environ.* 568, 327–338.
- Lam, S.K., Suter, H., Mosier, A.R., Chen, D., 2017. Using nitrification inhibitors to mitigate agricultural N_2O emission: a double-edged sword? *Glob. Chang. Biol.* 23, 485–489.
- Lamb, A., Green, R., Bateman, I., Broadmeadow, M., Bruce, T., Burney, J., Carey, P., Chadwick, D., Crane, E., Field, R., Goulding, K., Griffiths, H., Hastings, A., Kasoar, T., Kindred, D., Phalan, B., Pickett, J., Smith, P., Wall, E., zu Ermgassen, E.K.H.J., Balmford, A., 2016. Land sparing as a mechanism to offset greenhouse gas emissions from agriculture. *Nat. Clim. Change* 6 (5), 488–492.
- Laubach, J., Taghizadeh-Toosi, A., Gibbs, S.J., Sherlock, R.R., Kelliher, F.M., Grover, S.P.P., 2013. Ammonia emissions from cattle urine and dung excreted on pasture. *Biogeosciences* 10, 327–338.
- Ledgard, S.F., Menneer, J.C., Dexter, M.M., Kear, M.J., Lindsey, S., Peters, J.S., Pacheco, D., 2008. A novel concept to reduce nitrogen losses from grazed pastures by administering soil nitrogen process inhibitors to ruminant animals: a study with sheep. *Agric. Ecosyst. Environ.* 125, 148–158.
- Lockyer, D.R., Whitehead, D.C., 1990. Volatilization of ammonia from cattle urine applied to grassland. *Soil Biol. Biochem.* 22, 1137–1142.
- Luo, J., Ledgard, S., Wise, B., Welten, B., Lindsey, S., Judge, A., Sprosen, M., 2015. Effect of dicyandiamide (DCD) delivery method, application rate, and season on pasture urine patch nitrous oxide emissions. *Biol. Fertil. Soils* 51, 453–464.
- Luo, J., Balvert, S.F., Wise, B., Welten, B., Ledgard, S.F., de Klein, C.A.M., Lindsey, S., Judge, A., 2018. Using alternative forage species to reduce emissions of the greenhouse gas nitrous oxide from cattle urine deposited onto soil. *Sci. Total Environ.* 610–611, 1271–1280.
- Marsden, K.A., Scowen, M., Hill, P.W., Jones, D.L., Chadwick, D.R., 2015. Plant acquisition and metabolism of the synthetic nitrification inhibitor dicyandiamide and naturally-occurring guanidine from agricultural soils. *Plant Soil* 395, 201–214.
- Marsden, K.A., Jones, D.L., Chadwick, D.R., 2016. Disentangling the effect of sheep urine patch size and N loading rate on cumulative N_2O emissions. *Anim. Prod. Sci.* 56, 265–275.
- McGeough, K.L., Watson, C.J., Müller, C., Laughlin, R.J., Chadwick, D.R., 2016. Evidence that the efficacy of the nitrification inhibitor dicyandiamide (DCD) is affected by soil properties in UK soils. *Soil Biol. Biochem.* 94, 222–232.
- McGeough, K.L., McNeill, G., Laughlin, R.J., Watson, C.J., 2017. Agricultural Greenhouse Gas Inventory Research Platform - *InveN2Ory*. Dung and urine experimental site in County Down, 2012. Version:1. [dataset] Freshwater Biological Association [publisher]. <https://doi.org/10.17865/ghgn0570>.
- Minet, E.P., Ledgard, S.F., Lanigan, G.J., Murphy, J.B., Grant, J., Hennessy, D., Lewis, E., Forrester, P., Richards, K.G., 2016. Mixing dicyandiamide (DCD) with supplementary feeds for cattle: an effective method to deliver a nitrification inhibitor in urine patches. *Agric. Ecosyst. Environ.* 231, 114–121.
- Minet, E.P., Ledgard, S.F., Grant, J., Murphy, J.B., Krol, D.J., Lanigan, G.J., Luo, J., Richards, K.G., 2018. Feeding dicyandiamide (DCD) to cattle: an effective method to reduce N_2O emissions from urine patches in a heavy-textured soil under temperate climatic conditions. *Sci. Total Environ.* 615, 1319–1331.
- Misselbrook, T.H., Cardenas, L.M., Camp, V., Thorman, R.E., Williams, J.R., Rollett, A.J., Chambers, B.J., 2014. An assessment of nitrification inhibitors to reduce nitrous oxide emissions from UK agriculture. *Environ. Res. Lett.* 9. <https://doi.org/10.1088/1748-9326/9/11/115006>.
- Misselbrook, T.M., Fleming, H., Camp, V., Umstatter, C., Duthie, C.-A., Nicoll, L., Waterhouse, T., 2016. Automated monitoring of urination events from grazing cattle. *Agric. Ecosyst. Environ.* 230, 191–198.
- Monaghan, R.M., de Klein, C.A.M., 2014. Integration of measures to mitigate reactive nitrogen losses to the environment from grazed pastoral dairy systems. *J. Agric. Sci.* 152, 45–56.
- Pal, P., McMillan, A.M., Saggart, S., 2016. Pathways of dicyandiamide uptake in pasture plants: a laboratory study. *Biol. Fertil. Soils* 52, 539–546.
- Qui, W.H., Di, H.J., Cameron, K.C., Hu, C.X., 2010. Nitrous oxide emissions from animal urine as affected by season and a nitrification inhibitor dicyandiamide. *J. Soils Sediments* 10, 1229–1235.
- Reed, K.F., Moraes, L.E., Casper, D.P., Kebreab, E., 2015. Predicting nitrogen excretion from cattle. *J. Dairy Sci.* 98, 3025–3035.
- Reidy, B., Dammgen, U., Dohler, H., Eurich-Menden, B., van Evert, F.K., Hutchings, N.J., Luesink, H.H., Menzi, H., Misselbrook, T.H., Monteny, G.J., Webb, J., 2008. Comparison of models used for national agricultural ammonia emission inventories in Europe: liquid manure systems. *Atmos. Environ.* 42, 3452–3464.
- Selbie, D.R., Cameron, K.C., Di, H.J., Moir, J.L., Lanigan, G.J., Richards, K.G., 2014. The effect of urinary nitrogen loading rate and a nitrification inhibitor on nitrous oxide emissions from a temperate grassland soil. *J. Agric. Sci.* 152 (S1), 159–171.
- Selbie, D.R., Buckthought, L.E., Shepherd, M.A., 2015. Chapter four—the challenge of the urine patch for managing nitrogen in grazed pasture systems. *Adv. Agron.* 129, 229–292.
- Singh, U., Sanabria, J., Austin, E.R., Agyin-Birikorang, S., 2011. Nitrogen transformation, ammonia volatilization loss, and nitrate leaching in organically enhanced nitrogen fertilizers relative to urea. *Soil Sci. Soc. Am. J.* 76, 1842–1854.
- Sugimoto, Y., Ball, P.R., 1989. Nitrogen losses from cattle dung. In: Desroches, R. (Ed.), *Proceedings XVI International Grassland Congress. Vol. I*, pp. 153–154 Nice.
- Thorman, R.E., Nicholson, F.A., McMillan, S., Shrobsree, A., Smith, K.E., Williams, J.R., 2017a. Agricultural Greenhouse Gas Inventory Research Platform - *InveN2Ory*. Dung and urine experimental site in Warwickshire, 2013. Version:1. [dataset] Freshwater Biological Association [publisher]. <https://doi.org/10.17865/ghgn0652>.
- Thorman, R.E., Lathwood, T., Nicholson, F.A., Shrobsree, A., Smith, K.E., Williams, J.R., 2017b. Agricultural Greenhouse Gas Inventory Research Platform - *InveN2Ory*. Dung and urine experimental site in Ceredigion, 2013. Version:1. [dataset] Freshwater Biological Association [publisher]. <https://doi.org/10.17865/ghgn0644>.
- UNFCCC, 2016. National greenhouse gas emission submissions for the year 2013. http://unfccc.int/national_reports/annex_i_ghg_inventories/national_inventories_submissions/items/8812.php, Accessed date: 1 August 2016.
- van der Weerden, T.J., Styles, T.M., Rutherford, A.J., de Klein, C.A.M., Dynes, R., 2017. Nitrous oxide emissions from cattle urine deposited onto soil supporting a winter forage kale crop. *N. Z. J. Agric. Res.* 60, 119–130.
- van Groenigen, J.W., Kuikman, P.J., de Groot, W.J., Velthof, G.L., 2005. Nitrous oxide emission from urine-treated soil as influenced by urine composition and soil physical conditions. *Soil Biol. Biochem.* 37, 463–473.
- VSN International, 2015. *Genstat for Windows 18th Edition*. VSN International, Hemel Hempstead, UK Web page: Genstat.co.uk.
- Webb, J., Misselbrook, T.H., 2004. A mass-flow model of ammonia emissions from UK livestock production. *Atmos. Environ.* 38, 2163–2176.
- Welten, B.G., Ledgard, S.F., Luo, J., 2014. Administration of dicyandiamide to dairy cows via drinking water reduces nitrogen losses from grazed pastures. *J. Agric. Sci.* 152, 150–158.